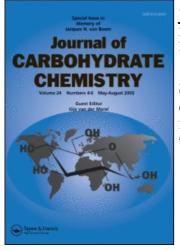
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SYNTHETIC STUDIES ON SIALOGLYCOCONJUGATES 54: SYNTHESIS OF I-ACTIVE GANGLIOSIDE ANALOG

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ABSTRACT

A stereocontrolled synthesis of I-active ganglioside analog is described. Glycosylation of 2-(trimethylsilyl)ethyl O-(2-O-benzyl-4,6-O-benzylidene- β -Dgalactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-benzyl- β -D-glucopyranoside (5) with methyl 4-Oacetyl-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio-B-D-glucopyranoside (10) by use of N-iodosuccinimide (NIS)-trifluoromethanesulfonic acid (TfOH) gave the desired trisaccharide 11, which was transformed into trisaccharide acceptor 14 via removal of the phthaloyl group followed by N-acetylation, and debenzylidenation. Glycosylation of 14 with methyl 3-O-benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido-1-thio-β-Dglucopyranoside (8) gave the biantennary compound 15, which was transformed into the acceptor 16. Dimethyl(methylthio)sulfonium triflate (DMTST)-promoted coupling of 16 with methyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-Dgalacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -2,4,6-tri-O-benzoyl-1-thio- β -D-galactopyranoside (17) afforded the desired hexasaccharide 19. Coupling of the hexasaccharide acceptor 20, prepared from 19 by reductive ring-opening of benzylidene acetal, with 17 gave octasaccharide derivative 21. Compound 21 was transformed, via removal of the benzyl group followed by O-acetylation, selective removal of the 2-(trimethylsilyl)ethyl group and subsequent imidate formation, into the final glycosyl donor 24. Condensation of 24 with (2S, 3R, 4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (18) gave the β glycoside 25, which on channeling through selective reduction of azido group, coupling of the amino group with octadecanoic acid, O-deacylation and saponification of the methyl ester group, gave the title compound 28.

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INTRODUCTION

Numerous glycosphingolipids are found as constituents of the outer surface of cell membranes.^{1,2} These compounds have important biological roles in such as cell growth, differentiation, adhesion, oncogenesis, receptor functions for viruses and bacterial toxins, and ligand activities for E-, P-, and L-selectins.² In order to investigate the functions of sialoglycoconjugates at the molecular level, we have synthesized a series of gangliosides and their analogs,³ by using our newly developed methods for ganglioside synthesis.⁴⁻⁶

In the studies on the isolation and characterization of various glycoconjugates from human lung, I- and sialyl I-antigen 1 have been recognized as specific markers for differentiated type cells in the developing lung of human embryos and in lung cancers.⁷ In order to elucidate the role of the biantennary structure in the function of the sialyl Iantigen, here we describe the synthesis of I-active ganglioside analog, VI³ Neu5Ac α , IV³ Neu5Ac α , II⁶ klado Lc₆ Cer, in which the lactotriose residue in crude sialyl Iantigen 1 is replaced by D-glucopyranose.

RESULTS AND DISCUSSION

For the synthesis of desired I-active ganglioside analog, we have selected 2-(trimethylsilyl)ethyl *O*-(2-acetamido-3-*O*-benzyl-4,6-*O*-benzylidene-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)-*O*-[(2-acetamido-3,6-di-*O*-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)]-*O*-(2-*O*-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-glucopyranoside (16) as a key glycosyl acceptor, which has the biantennary structure obtained by coupling of the glucosamine derivative 8 and the lactotriose acceptor 14 constructed from the lactose unit 5 and the glucosamine unit 10. We have employed methyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-2,4,6-tri-*O*-benzoyl-1-thio- β -D-galactopyranoside⁸ (17) as the glycosyl donor.

2-(Trimethylsilyl)ethyl O-(2-O-benzyl-4,6-O-benzylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (5) was obtained in good yield from 2-(trimethylsilyl)ethyl O-[3-O-(4-methoxybenzyl)- β -D-galactopyranosyl]-(1 \rightarrow 4)- β -D-glucopyranoside⁸ (2) via 4,6-O-benzylidenation, O-benzylation, and oxidative removal of the 4-methoxybenzyl group with 2,3-dichloro-5,6-dicyanobenzoquinone⁹. One of the glucosamine donors, methyl 3-O-benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido-1thio- β -D-glucopyranoside (8), was prepared from methyl 2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside¹⁰ (6) by 4,6-O-benzylidenation and 3-O-benzylation. The glycosyl donor 10 was obtained from 8 by the reductive ring-opening¹¹ of benzylidene acetal with sodium cyanoborohydride-hydrogen chloride according to the method of Garegg *et al.* and subsequent acetylation. The glycosylation of 5 with 10 in dichloromethane for 2 h at 0 °C in the presence of *N*-iodosuccinimide (NIS)-trifluoromethanesulfonic acid (TfOH) and powdered molecular sieves 4Å (MS-4Å) afforded the desired β -glycoside 11 in 94% yield. The observed chemical shifts and coupling constants of the glucosamine unit for H-1 (δ 5.06, J_{1,2} = 11.2 Hz) and H-4 (δ 5.12, J_{3,4} = J_{4,5} = 9.7 Hz) indicated the structure assigned. *O*-Deacetylation and conversion from the phthalimide to the acetamide by heating with hydrazine hydrate in aqueous 95% ethanol, *N*-acetylation, and *O*-acetylation of 11 afforded the lactotriose derivative 13. The hydrolysis of the benzylidene group in 13 gave the lactotriose acceptor 14.

The condensation of 14 with 8 in a similar condition as described for the glycosylation of 5 with 10 gave the biantennary compound 15 in 61% yield. Significant signals of the glucosamine unit in the ¹H NMR spectrum of 15 were a one-proton doublet at δ 5.45 (J_{1,2} = 10.8 Hz, H-1), a one-proton singlet at 5.60 (PhCH), and forty-four aromatic protons at δ 7.17-7.67 (9 Ph), indicating the newly formed glycosidic linkage to be β . By removal of the phthaloyl, followed by *N*-acetylation, the branching tetrasaccharide acceptor 16 was formed from 15 in 89% yield.

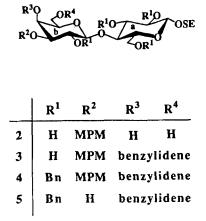
The glycosylation of 16 with 17 in the presence of dimethyl(methylthio)sulfonium triflate⁴ (DMTST) as the glycosyl promoter and powdered MS-4Å in dichloromethane for 16 h at 0~6 °C gave the hexasaccharide 19 in 74% yield, which had the expected stereochemistry. The ¹H NMR spectrum of 19 showed the presence of seven three-proton singlets at δ 1.49, 1.78, 1.92-2.15 (*O*-acetyl and *N*-acetyl), a three-proton singlet at δ 3.83 (*O*-methyl), fifty-five aromatic protons (11Ph) at δ 7.13-8.30, and a one-proton doublet of doublets at δ 5.51 (J_{1,2} = 8.4 Hz, J_{2,3} = 12.6 Hz, H-2d), and those data are consistent with the structure assigned. The reductive ring-opening of the benzylidene acetal in 19 afforded the hexasaccharide acceptor 20 in 80% yield.

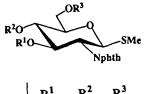
The glycosylation of 20 with 17 by use of DMTST in the presence of powdered MS-4Å in dichloromethane for 16 h at 0~6 °C gave expected octasaccharide derivative 21 in 28% yield; significant signals of 21 in ¹H NMR spectrum were a two-proton multiplet at δ 2.45 (H-3e-*eq* and H-3h-*eq*), two three-proton singlets at δ 3.78, 3.82 (*O*-methyl), seventy aromatic protons (14Ph) at δ 7.03-8.31, and a two-proton doublet of doublets at δ 5.52 (J_{1,2} = 7.5 Hz, J_{2,3} = 13.3 Hz, H-2d and H-2g), indicating the structure assigned.

Catalytic hydrogenolysis (10% Pd-C) of the benzyl group in 21 in ethanol-acetic acid for 48 h at 45 °C and subsequent O-acetylation gave the per-O-acyl compound 22 in 31% yield. Treatment¹² of 22 with trifluoroacetic acid in dichloromethane for 30 min at

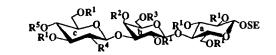
$$\label{eq:gamma} \begin{array}{l} \operatorname{Neu5Ac}(2 \to 3) Gal\beta(1 \to 4) GlcNAc\beta(1 \to 6) \\ Gal\beta(1 \to 4) GlcNAc\beta(1 \to 3) Gal\beta(1 \to 4) Glc\beta(1 \to 1) Cer \\ \operatorname{Neu5Ac}(2 \to 3) Gal\beta(1 \to 4) GlcNAc\beta(1 \to 3) \end{array}$$

1

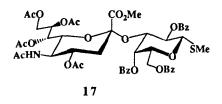


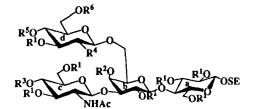


	R'	<u></u>	_ <u>R°</u>
6	H	н	H
7	н	benzy	ylidene
8	Bn	benzy	ylidene
9	Bn	Н	Bn
10	Bn	Ac	Bn

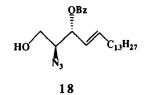


	R ¹	R ²	R ³	R ⁴	R ⁵
11	Bn	benzy	lidene	Nphth	Ac
12	Bn	benzy	lidene	NHAc	н
13	Bn	benzy	lidene	NHAc	Ac
14	Bn	н	Н	NHAc	Ac



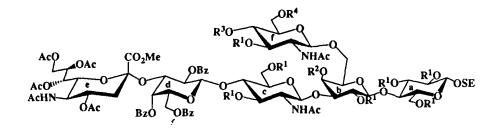


	R ¹		R ³	R ⁴	R ⁵	R ⁶
15	Bn	Н	Ac	Nphth NHAc	benzy	lidene
16	Bn	H	Н	NHAc	benzy	lidene

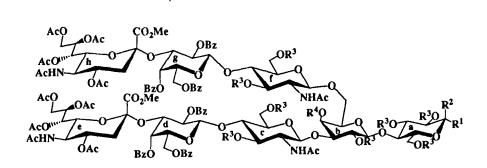


SE = 2-(trimethylsilyl)ethyl Bn = benzyl Bz = benzoyl MPM = p-methoxybenzyl

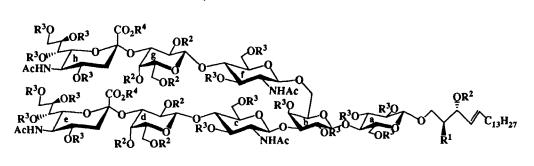
.



		R ¹	R ²	R ³	R ⁴
1	9	Bn	н	benzy	lidene
2	0	Bn	Н	Н	Bn



	R ¹	R ²	R ³	R ⁴
21	OSE	Н	Bn	Н
22	OSE	Н	Ac	Ac
23 24	н , он		Ac	Ac
24	H OC(=NH)CCl ₃		Ac	Ac



	R ¹	R ²	<u>R³</u>	R ⁴
25	N ₃	Βz	Ac	Me
26	NH ₂	Βz	Ac	Me
27	NHCO(CH ₂) ₁₆ CH ₃	Βz	Ac	Me
28	NHCO(CH ₂) ₁₆ CH ₃ NHCO(CH ₂) ₁₆ CH ₃	Н	н	Н

0 °C gave the 1-hydroxy compound 23. When treated with trichloroacetonitrile in dichloromethane in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) for 3 h at 0 °C, 23 gave the trichloroacetimidate 24 in 59% yield. The ¹H NMR data for Gal unit in 24 [δ 6.44 (J_{1,2} = 3.8 Hz, H-1a), 8.61 (C=NH)] indicated the trichloroacetimidate to be α .

The final glycosylation of (2S,3R,4E)-2-azido-3-*O*-benzoyl-4-octadecene-1,3-diol¹³ (18) with 24 in dichloromethane in the presence of boron trifluoride etherate^{13b,14} for 8 h at 0 °C afforded the desired β -glycoside 25 in 46% yield. Selective reduction¹⁵ of azido group in 25 with hydrogen sulfide in aq 83% pyridine for 3 days at 0 °C gave the amine, which, on condensation with octadecanoic acid, using 1-ethyl-3-(3dimethylaminopropyl)carbodiimide (WSC) in dichloromethane gave the acylated sialyl Iantigen analog 27 in 62% yield, after chromatography.

Finally, O-deacylation of 27 with sodium methoxide in methanol, with subsequent saponification of the methyl ester group, yielded the desired sialyl I-antigen analog 28, α -Neu5Ac-(2 \rightarrow 3)- β -D-Gal-(1 \rightarrow 4)- β -D-GlcNAc-(1 \rightarrow 6)-[α -Neu5Ac-(2 \rightarrow 3)- β -D-Gal-(1 \rightarrow 4)- β -D-GlcNAc-(1 \rightarrow 4)- β -D-GlcNAc-(1 \rightarrow 4)- β -D-Glc-(1 \rightarrow 1)-Cer, in quantitative yield after chromatography on a column of Sephadex LH-20.

EXPERIMENTAL

General Procedures. Specific rotations were determined with a Union PM-201 polarimeter at 25 °C. ¹H NMR spectra were recorded with a JEOL JNM-GX 270 spectrometer. Preparative chromatography was performed on silica gel (Wako Chemical Co., 200 mesh) with the solvent systems specified. Concentrations were conducted *in vacuo*.

2-(Trimethylsilyl)ethyl O-[4,6-O-Benzylidene-3-O-(4-methoxybenzyl)- β -D-galactopyranosyl]-(1 \rightarrow 4)- β -D-glucopyranoside (3). To a solution of 2-(trimethylsilyl)ethyl O-[3-O-(4-methoxybenzyl)- β -D-galactopyranosyl]-(1 \rightarrow 4)- β -Dglucopyranoside⁸ (2; 3.5 g, 6.20 mmol) in DMF (50 mL) was added Drierite (5.0 g), and the mixture was stirred for 2 h at room temperature. Benzaldehyde dimethyl acetal (1.9 mL, 12.40 mmol) and *p*-toluenesulfonic acid monohydrate (50 mg) were added, and the mixture was stirred for 16 h at room temperature, then neutralised with Amberlite IR-410 (HO⁻) resin and concentrated. Column chromatography (3:1 AcOEt-hexane) of the residue on silica gel (100 g) afforded 3 (2.9 g, 72%) as an amorphous mass: [α]_D+16.5° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 0.98 (m, 2H, Me₃SiCH₂CH₂), 3.76 (s, 3H, CH₃OC₆H₄CH₂), 4.19, 4.63 (2d, 2H, J_{gem} = 12.3 Hz, CH₃OC₆H₄CH₂), 4.28, 4.43 (2d, 2H, J = 8.3 Hz, 8.1 Hz, H-1a, 1b), 5.32 (s, 1H, PhCH), and 6.82-7.48 (m, 9H, 2Ph).

Anal. Calcd for C₃₂H₄₆O₁₂Si (650.8): C, 59.06; H, 7.12. Found: C, 58.92; H, 7.29

2-(Trimethylsilyl)ethyl O-[2-O-Benzyl-4,6-O-benzylidene-3-O-(4-methoxybenzyl)- β -D-galactopyranosyl]-(1 \rightarrow 4)-2, 3, 6-tri-O-benzyl- β -D-glucopyranoside (4). To a solution of 3 (2.9 g, 4.46 mmol) in *N*,*N*-dimethyl-formamide (DMF, 25 mL) was added a suspension of sodium hydride in oil (1.1 g, 60% of sodium hydride by weight). The mixture was stirred for 30 min at 0 °C, benzyl bromide (3.2 mL, 26.8 mmol) was added dropwise, and stirring was continued for 3 h at room temperature. The reaction was monitored by TLC and, when complete, methanol (5 mL) was added, and the mixture was concentrated and extracted with dichloromethane. The extract was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (1:1 AcOEt-hexane) of the residue on silica gel (100 g) gave 4 (4.1 g, 88%) as a syrup: [α]_D +14.9° (*c* 1.1, CHCl₃); ¹H NMR (CDCl₃) δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 3.76 (s, 3H, CH₃OC₆H₄CH₂), 4.32, 4.51 (2d, 2H, J_{gem} = 12.3 Hz, CH₃OC₆H₄CH₂), 4.37, 4.43 (2d, 2H, J = 7.9 Hz, H-1a, 1b), 5.42 (s, 1H, PhCH), and 6.80-7.51 (m, 29H, 6Ph).

Anal. Calcd for C₆₀H₇₀O₁₂Si (1011.3): C, 71.26; H, 6.98. Found: C, 71.23; H, 6.68.

2-(Trimethylsilyl)ethyl O-(2-O-Benzyl-4,6-O-benzylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (5). To a stirred solution of 4 (7.6 g, 7.48 mmol) in dichloromethane (90 mL) were added 2,3dichloro-5,6-dicyanobenzoquinone (2.6 g, 11.2 mmol) and water (3 mL), and stirring was continued for 1 h at room temperature. The precipitate was collected and washed with dichloromethane, and the combined filtrate and washings were washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (1:5 AcOEt-hexane) of the residue on silica gel (300 g) gave 5 (5.4 g, 82%) as an amorphous mass: $[\alpha]_D$ -4.1° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 2.41 (broad d, 1H, OH), 5.50 (s, 1H, PhCH), and 7.15-7.48 (m, 25H, 5Ph).

Anal. Calcd for C₅₂H₆₂O₁₁Si (891.1): C, 70.09; H, 7.01. Found: C, 69.83; H, 6.97.

Methyl 4,6-O-Benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (7). To a solution of methyl 2-deoxy-2-phthalimido-1-thio- β -Dglucopyranoside (6; 10.4 g, 31 mmol) in DMF (100 mL) was added Drierite (10 g), and the mixture was stirred for 2 h at room temperature. Benzaldehyde dimethyl acetal (9.2 mL, 61 mmol) and *p*-toluenesulfonic acid monohydrate (90 mg) were added, and the mixture was stirred for 48 h at room temperature, then neutralised with Amberlite IR-410 (HO⁻) resin and concentrated. Column chromatography (3:2 AcOEt-hexane) of the residue on silica gel (300 g) afforded 7 (8.9 g, 68%) as an amorphous mass: $[\alpha]_D$ +48.2° (*c* 1.2, CHCl₃); ¹H NMR (CDCl₃) δ 2.18 (s, 3H, SMe), 3.62 (t, 1H, Jgem = J_{5,6'} = 8.8 Hz, H-6'), 3.73 (dt, 1H, J_{4,5} = 8.8 Hz, J_{5,6} = 4.2 Hz, H-5), 3.82 (t, 1H, H-6), 4.36 (t, 1H, J_{1,2} = J_{2,3} =10.3 Hz, H-2), 4.42 (dd, 1H, J_{3,4} = 9.7 Hz, H-4), 4.70 (dd, 1H, H-3) 5.31 (d, 1H, H-1), 5.58 (s, 1H, PhCH), and 7.25-8.04 (m, 9H, 2Ph).

Anal. Calcd for C₂₂H₂₁NO₆S (427.5): C, 61.81; H, 4.95; N, 3.28. Found: C, 61.61; H, 4.67; N, 3.30.

Methyl 3-O-Benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (8). To a solution of 7 (8.93 g, 20.9 mmol) in *N*,*N*dimethylformamide (DMF, 25 mL) was added a suspension of sodium hydride in oil (1.25 g, 60% of sodium hydride by weight). The mixture was stirred for 30 min at 0 °C, benzyl bromide (3.2 mL, 31.3 mmol) was added dropwise, and stirring was continued for 16 h at room temperature. The reaction was monitored by TLC and, when complete, methanol (5 mL) was added, and the mixture was worked up as described for 4. Column chromatography (1:1 AcOEt-hexane) of the residue on silica gel (100 g) gave 8 (7.6 g, 88%) as a syrup: $[\alpha]_D$ +54.1° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 2.14 (s, 3H, SMe), 3.71 (m, 1H, H-5), 3.77 (dd, 1H, Jgem = J_{5,6}' = 8.8 Hz, H-6'), 3.85 (dd, 1H, J_{5,6} = 5.7 Hz, H-6), 4.33 (t, 1H, J_{1,2} = J_{2,3} =10.1 Hz, H-2), 4.44 (dd, 1H, J_{3,4} = 10.3 Hz, H-4), 4.49 (dd, 1H, H-3) 5.24 (d, 1H, H-1), 5.64 (s, 1H, PhCH), and 6.85-7.85 (m, 14H, 3Ph).

Anal. Calcd for C₂₉H₂₇NO₆S(517.6): C, 67.29; H, 5.26; N, 2.71. Found: C, 67.38; H, 5.13; N, 2.70

Methyl 3,6-Di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (9). To a solution of 8 (4.0 g, 7.73 mmol) in dry tetrahydrofuran (120 mL) were added molecular sieves 3Å (MS-3Å; 20 g), and the mixture was stirred for 1 h at room temperature, and sodium cyanoborohydride (7.3 g) was gradually added. After the reagent had dissolved, hydrogen chloride in ether was added dropwise at room temperature until the evolution of gas ceased. TLC indicated that the reaction was complete after 5 min. The mixture was neutralized with triethylamine and filtered, the residue was washed with methanol and the combined filtrate and washings were concentrated then extracted with dichloromethane. The extract was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (1:3 AcOEt-hexane) of the residue on silica gel (100 g) afforded 9 (3.78 g, 94%) as an amorphous mass: [α]_D +47.9° (*c* 0.9, CHCl₃); ¹H NMR (CDCl₃) δ 2.10(s, 3H, SMe), 4.26 (t, 1H, J_{1,2} = J_{2,3} =9.9 Hz, H-2), 4.29 (dd, 1H, J_{3,4} = 7.4 Hz, J_{4,5} = 4.5 Hz, H-4), 5.17 (d, 1H, H-1), and 6.85-7.85 (m, 14H, 3Ph).

Anal. Calcd for C₂₉H₂₉NO₆S(519.6): C, 67.03; H, 5.63; N, 2.70. Found: C, 67.04; H, 5.74; N, 2.59.

Methyl 4-O-Acetyl-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (10). A solution of 9 (4.17 g, 8.0 mmol) in acetic anhydride (25 mL) and pyridine (40 mL) was stirred for 3 h at room temperature, and methanol (30 mL) was added. The solution was concentrated to a syrup which was extracted with dichloromethane. The extract was successively washed with 2 M hydrochloric acid and water, dried (Na₂SO₄) and concentrated. Column chromatography (1:3 AcOEt-hexane) of the residue on silica gel (200 g) gave 10 (4.50 g, quantitative) as an amorphous mass: [α]_D+112.4° (*c* 0.17, CHCl₃); ¹H NMR (CDCl₃) δ 1.93,2.10(2s, 6H, AcO and MeS), 4.37 (t, 1H, J_{1,2} = J_{2,3} =10.7 Hz, H-2), 4.51 (dd, 1H, J_{3,4} = 9.0 Hz, H-3), 5.18 (dd, 1H, J_{4,5} = 10.4 Hz, H-4), 5.20 (d, 1H, H-1), and 6.87-7.75 (m, 14H, 3Ph).

Anal. Calcd for C₃₁H₃₁NO₇S(561.7): C, 66.29; H, 5.56; N, 2.49. Found: C, 66.44; H, 5.67; N, 2.49

2-(Trimethylsilyl)ethyl O-(4-O-Acetyl-3,6-di-O-benzyl-2-deoxy-2phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 3)-O-(2-O-benzyl-4,6-O-benzylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (11). To a solution of 5 (200 mg, 0.22 mmol) and 10 (214 mg, 0.38 mmol) in dry dichloromethane (5 mL) was added MS-4Å (0.5 g), and the mixture was stirred for 5 h at room temperature then cooled to 0 °C. To the cooled mixture were added, with stirring, N-iodosuccinimide (NIS; 172 mg, 0.76 mmol) and trifluoromethanesulfonic acid (TfOH; 17 μ L, 0.08 mmol), and the stirring was continued for 2 h at 0 °C. The precipitate was filtered off, and washed thoroughly with dichloromethane. The filtrate and washings were combined, and the solution was successively washed with M Na₂CO₃, M Na₂S₂O₃ and water, dried (Na₂SO₄) and concentrated. Column chromatography (1:3 AcOEt-hexane) of the residue on silica gel (20 g) gave 11 (296 mg, 94%) as an amorphous mass: $[\alpha]_D$ +120.6° (c 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 0.96 (m, 2H, Me₃SiCH₂CH₂), 2.01 (s, 3H, AcO), 5.06 (d, 1H, J_{1,2} = 11.2 Hz, H-1c), 5.12 (t, 1H, J_{3,4} = J _{4,5} = 9.7 Hz, H-4c), 5.41 (s, 1H, PhCH), and 6.78-7.51 (m, 39H, 8Ph)

Anal. Calcd for C₈₂H₈₉NO₁₈Si (1404.7): C, 70.12; H, 6.39; N, 1.00. Found: C, 70.02; H, 6.55; N, 1.01.

2-(Trimethylsilyl)ethyl O-(2-Acetamido-4-O-acetyl-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-O-(2-O-benzyl-4,6-O-benzylidene- β -

D-galactopyranosyl)- $(1 \rightarrow 4$)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (13). A solution of 11 (1.82 g, 1.30 mmol) in aqueous 95% ethanol (50 mL) was treated with hydrazine hydrate (0.6 mL) for 12 h under reflux. The precipitate was collected and washed with ethanol, and the combined filtrate and washings were concentrated. The residue was treated with acetic anhydride (1 mL) and pyridine (2 mL) for 2 h at room temperature, and methanol (2 mL) was added. The mixture was concentrated, and a solution of the residue in dichloromethane was successively washed with 2 M hydrochloric acid, water, and M sodium carbonate, dried (Na₂SO₄) and concentrated. Column chromatography (1:3 AcOEt-hexane) of the residue on silica gel (100 g) afforded 7 (1.58 g, 92%) as an amorphous mass: $[\alpha]_D$ +128.1° (c 0.9, CHCl₃); ¹H NMR (CDCl₃) 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.40, 2.01 (2s, 6H, AcO and AcN), 5.13 (t, 1H, J_{3,4} = J_{4,5} = 8.2 Hz, H-4c), 5.42 (s, 1H, PhCH), and 7.16-7.55 (m, 35H, 7Ph)

Anal. Calcd for C₆₈H₈₅NO₁₇Si (1216.5): C, 67.14; H, 7.04; N, 1.15. Found: C, 67.38; H, 7.10; N, 1.11.

2-(Trimethylsilyl)ethyl O-(2-Acetamido-4-O-acetyl-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-O-(2-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (14). A solution of 13 (1.58 g, 1.20 mmol) in aqueous 80% acetic acid (30 mL) was heated for 24 h at 60 °C and concentrated. Column chromatography (1:1 AcOEt-hexane) of the residue on silica gel (100 g) gave 14 (1.35 g, 92%) as an amorphous mass: [α]_D +174.0° (c 0.80, CHCl₃); ¹H NMR (CDCl₃) δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 1.42, 1.89 (2s, 6H, AcO and AcN), 5.08 (d, J_{1,2} = 8.3 Hz, H-1c), 5.13 (t, 1H, J_{3,4} = J 4,5 = 8.2 Hz, H-4c), and 7.16-7.55 (m, 30H, 6Ph).

Anal. Calcd for C₆₁H₈₁NO₁₇Si (1128.4): C, 64.93; H, 7.24; N, 1.24. Found: C, 64.79; H, 7.41; N, 1.06.

2-(Trimethylsilyl)ethyl O-(2-Acetamido-4-O-acetyl-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-O-[(3-O-benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 6)]-O-(2-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (15). To a solution of 14 (1.05 g, 0.85 mmol) and 8 (0.75 g, 1.45 mmol) in dry dichloromethane (20 mL) was added MS-4Å (3.0 g), and the mixture was stirred for 5 h at room temperature then cooled to 0 °C. To the cooled mixture were added, with stirring, NIS (654 mg, 2.91 mmol) and TfOH (66 μ L, 0.34 mmol), and the stirring was continued for 2 h at 0 °C, and then worked-up, as described for 11. Column chromatography (1:3 AcOEt-hexane) of the residue on silica gel (20 g) gave 15 (890 mg, 61%) as an amorphous mass: $[\alpha]_D$ +42.0° (c 1.6, CHCl₃); ¹H NMR (CDCl₃) δ 1.03 (m, 2H, Me₃SiCH₂CH₂), 1.70, 1.85 (2s, 6H, AcO and AcN), 5.11 (t, 1H, $J_{3,4} = J_{4,5} = 8.6$ Hz, H-4c) 5.16 (d, 1H, $J_{1,2} = 10.8$ Hz, H-1c), 5.45 (d, 1H, $J_{1,2} = 10.8$ Hz, H-1d), 5.60 (s, 1H, PhCH) and 7.17-7.67 (m, 44H, 9Ph)

Anal. Calcd for C₉₉H₁₁₂N₂O₂₄Si (1742.1): C, 68.26; H, 6.48; N, 1.61. Found: C, 68.23; H, 6.18; N, 1.52.

2-(Trimethylsilyl)ethyl O-(2-Acetamido-3,6-di-O-benzyl-2-deoxy-β-D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-[(2-acetamido-3-O-benzyl-4,6-O-benzylidene-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 6)$]-O-(2-O-benzyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -2,3,6-tri-O-benzyl- β -D-glucopyranoside (16). A solution of 15 (810 mg, 0.47 mmol) in aqueous 95% ethanol (20 mL) was treated with hydrazine hydrate (1.0 mL) for 72 h under reflux. The precipitate was collected and washed with ethanol, and the combined filtrate and washings were concentrated. The residue was treated with acetic anhydride (1 mL) in methanol (10 mL) for 2 h at room temperature, pyridine (2 mL) was added, the mixture was concentrated, and a solution of the residue in dichloromethane was successively washed with 2 M hydrochloric acid, water, and M sodium carbonate, dried (Na2SO4) and concentrated. Column chromatography (2:1 AcOEt-hexane) of the residue on silica gel (100 g) afforded 16 (664 mg, 89%) as an amorphous mass: [α]_D +11.9° (c 1.2, CHCl₃); ¹H NMR (CDCl₃) δ 1.00 (m, 2H, Me₃SiCH₂CH₂), 1.92, 1.95 (2s, 6H, 2AcN), 5.18(d, 1H, J_{1,2} = 9.2 Hz, H-1c), 5.32(d, 1H, J_{1,2} = 8.1 Hz, H-1d), 5.56 (s, 1H, PhCH) and 7.01-7.04 (m, 40H, 8Ph)

Anal. Calcd for C₉₃H₁₁₂N₂O₂₃Si (1654.0): C, 67.53; H, 6.83; N, 1.69. Found: C, 67.28; H, 6.79; N, 1.98.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4, 7, 8, 9-tetra-Oacetyl-3, 5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2- \rightarrow 3)-O-(2, 4, 6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3, 6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-[O-(2acetamido-3-O-benzyl-4,6-O-benzylidene-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)]-O-(2-O-benzyl-4,6-O-benzylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (19). To a solution of 16 (410 mg, 0.262 mmol) and methyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-Dglycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-2,4,6-tri-O-benzoyl-1-thio- β -Dgalactopyranoside⁸ (17: 521 mg,0.523 mmol) in dry dichloromethane (25 mL) was added MS-4Å (7.5 g), and the mixture was stirred for 5 h at room temperature and cooled to 0 °C. Dimethyl(methylthio)sulfonium triflate (DMTST; 540 mg, 2.09 mmol) was added to the mixture, and the mixture was stirred for 16 h at 6 °C, filtered, washed with dichloromethane. The combined filtrate and washings were washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (4:1 AcOEt-hexane) of the residue on silica gel (10 g) gave **19** (490 mg, 74%) as an amorphous mass: $[\alpha]_D$ +5.28° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 0.99 (m, 2H, Me₃SiCH₂CH₂), 1.49, 1.78, 1.92, 1.98, 2.15 (5s, 21H, 4AcO and 3AcN), 2.47 (dd, 1H, Jgem = 13.1 Hz, J_{3eq,4} = 4.9 Hz, H-3e-eq), 3.83 (s, 3H, MeO), 5.10 (d, 1H, J_{1,2} = 12.9 Hz, H-1c), 5.21 (d, 1H, J_{1,2} = 12.5 Hz, H-1f), 5.51 (dd, 1H, J_{1,2} = 8.4 Hz, J_{2,3} = 12.6 Hz, H-2d), 5.57 (s, 1H, PhCH), 5.71 (m, 1H, H-8e), and 7.13-8.30 (m, 55H, 11Ph).

Anal. Calcd for C₁₃₆H₁₆₉N₃O₄₁Si (2529.9): C, 64.57; H, 6.73, N, 1.66. Found: C, 64.37; H, 6.45; N, 1.68.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4, 7, 8, 9-tetra-Oacetyl-3, 5-dideoxy-D-glycero-a-D-galacto-2-nonulopyranosylonate)-(2- \rightarrow 3)-O-(2, 4, 6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3, 6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -[O-(2acetamido-3, 6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 6)$]-O-(2-O-benzyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -2, 3, 6-tri-O-benzyl- β -D-glucopyranoside (20). To a solution of 19 (92 mg, 0.037 mmol) in dry tetrahydrofuran (3 mL) were added MS-3Å (0.8 g), and the mixture was stirred for 1 h at room temperature, and sodium cyanoborohydride (35 mg) was gradually added. After the reagent had dissolved, hydrogen chloride in ether was added dropwise at room temperature until the evolution of gas ceased. TLC indicated that the reaction was complete after 5 min, and then worked-up, as described for 9. Column chromatography (3:1 AcOEt-hexane) of the residue on silica gel (10 g) afforded 20 (74 mg, 80%) as an amorphous mass: $[\alpha]_D$ +3.0° (c 2.2, CHCl₃); ¹H NMR (CDCl₃) δ 0.96 (m, 2H, Me₃SiCH₂CH₂), 1.49, 1.78-2.00 (7s, 21H, 4AcO and 3AcN), 2.47 (m, 1H, H-3e-eq), 3.83 (s, 3H, MeO), 5.70 (m, 1H, H-8e), and 7.14-8.26 (m, 55H,11Ph)

Anal. Calcd for C₁₃₆H₁₇₁N₃O₄₁Si (2531.9): C, 64.52; H, 6.81, N, 1.66. Found: C, 64.53; H, 6.92; N, 1.55.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4, 7, 8, 9-tetra-Oacetyl-3, 5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2- \rightarrow 3)-O-(2, 4, 6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-[(methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2nonulopyranosylonate)-(2 \rightarrow 3)-O-(2, 4, 6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)]-O-(2-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O- **benzyl**-β-**D**-glucopyranoside (21). To a solution of 20 (73 mg, 0.029 mmol) and 17 (58 mg, 0.058 mmol) in dry dichloromethane (0.8 mL) was added MS-4Å (1.2 g), and the mixture was stirred for 5 h at room temperature and cooled to 0 °C. DMTST (60 mg, 0.232 mmol) was added to the mixture, the mixture was stirred for 16 h at 6 °C, and then worked-up, as described for 19, affording 21 (28 mg, 28%) as an amorphous mass: $[\alpha]_D$ +14.0° (*c* 2.26, CHCl₃); ¹H NMR (CDCl₃) δ 1.00 (m, 2H, Me₃SiCH₂CH₂), 1.50, 1.79-2.19 (9s, 36H, 8AcO and 4AcN), 2.45 (m, 2H, H-3e-*eq* and H-3h-*eq*), 3.78, 3.82 (2s, 6H, 2MeO), 5.07 (d, 1H, J_{1,2} = 9.5 Hz, H-1c), 5.24 (d, 1H, J_{1,2} = 7.7 Hz, H-1f), 5.52 (dd, 2H, J_{1,2} = 7.5 Hz, J_{2,3} = 13.3 Hz, H-2d and H-2g), and 7.03-8.31 (m, 70H, 14Ph).

Anal. Calcd for C₁₈₃H₂₀₄N₄O₆₀Si (3447.7): C, 63.75; H, 5.96, N, 1.63. Found: C, 63.90; H, 6.07; N, 1.63.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4, 7, 8, 9-tetra-Oacetyl-3, 5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2- \rightarrow 3)-O-(2, 4, 6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -[(methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-a-D-galacto-2nonulopyranosylonate)- $(2 \rightarrow 3)$ -O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-3, 6-di-O-acetyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 6)$]-O-(2,4-di-O-acetyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -2,3,6tri-O-acetyl-β-D-glucopyranoside (22). A solution of 21 (226 mg, 0.065 mmol) in ethanol (42 mL) and acetic acid (5.7 mL) was hydrogenated in the presence of 10% Pd-C (300 mg) for 48 h at 45 °C, then filtered and concentrated. The residue was acetylated with acetic anhydride (0.3 mL)-pyridine (1.2 mL) for 16 h at room temperature. The product was purified by chromatography on a column of silica gel (20 g) with 4:1 AcOEt-hexane afforded 22 (64 mg, 31%) as an amorphous mass: $[\alpha]_D$ +3.47° (c 1.3, CHCl₃); ¹H NMR (CDCl₃) δ 0.94 (m, 2H, Me₃SiCH₂CH₂), 1.55, 1.57, 1.68-2.19 (14s, 63H, 17AcO and 4AcN), 2.49 (m, 2H, H-3e-eq and H-3h-eq), 3.82, 3.84 (2s, 6H, 2MeO), 5.64 (m, 2H, H-8e and H-8h), and 7.21-8.22 (m, 30H, 6Ph).

Anal. Calcd for C₁₄₅H₁₇₄N₄O₆₉Si (3105.0): C, 56.09; H, 5.65, N, 1.80. Found: C, 55.95; H, 5.82; N, 1.62.

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glyc $ero-\alpha-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-benzoyl \beta-D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3,6-di-O-acetyl-2-deoxy-\beta D-glucopyranosyl)-(1 \rightarrow 3)-O-[(methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl 3,5-dideoxy-D-glycero-\alpha-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-$ (2,4,6-tri-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-*O*-(2-acetamido-3,6di-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)]-*O*-(2,4-di-*O*-acetyl- β -Dgalactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-acetyl-D-glucopyranose (23). To a solution of 22 (52 mg, 0.017 mmol) in dichloromethane (1 mL) was added trifluoroacetic acid (3 mL) at 0 °C, and the mixture was stirred for 30 min at 0 °C and concentrated. Column chromatography (20:1 CH₂Cl₂-MeOH) of the residue on silica gel (10 g) gave 23 (41 mg, 82%) as an amorphous mass: [α]_D +9.3° (*c* 0.8, CHCl₃); ¹H NMR (CDCl₃) δ 1.56, 1.59, 1.78-2.19 (9s, 63H, 17AcO and 4AcN), 2.44 (m, 2H, H-3e-*eq* and H-3h-*eq*), 3.78, 3.81 (2s, 6H, 2MeO), 5.62 (m, 2H, H-8e and H-8h), and 7.19-8.19 (m, 30H, 6Ph)

Anal. Calcd for C₁₄₀H₁₆₂N₄O₆₉ (3004.8): C, 55.96; H, 5.43, N, 1.86. Found: C, 55.82; H, 5.60; N, 1.68

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glyc $ero-\alpha-D-galacto-2$ -nonulopyranosylonate)- $(2\rightarrow 3)-O-(2,4,6-tri-O-benzoy)$ - β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-[(methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$ -O- $(2,4,6-\text{tri}-O-\text{benzoyl}-\beta-D-\text{galactopyranosyl})-(1\rightarrow 4)-O-(2-\text{acetamido}-3,6$ di-O-acetyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 6)$]-O-(2,4-di-O-acetyl- β -Dgalactopyranosyl)- $(1 \rightarrow 4)$ -2,3,6-tri-O-acetyl- α -D-glucopyranosyl Trichloroacetimidate (24). To a solution of 23 (40 mg, 0.013 mmol) in dichloromethane (1.0 mL) and trichloroacetonitrile (0.05 mL) was added 1,8diazabicyclo[5.4.0]undec-7-ene (DBU, 5.0 mg) at -5 °C, and the mixture was stirred for 3 h at 0 °C, then concentrated. Column chromatography (30:1 dichloromethanemethanol) of the residue on silica gel (10 g) gave 24 (24 mg, 59%) as an amorphous mass: $[\alpha]_D$ +11.4° (c 0.5, CHCl₃); ¹H NMR (CDCl₃) δ 1.53, 1.54, 1.78-2.21 (15s, 63H, 17AcO and 4AcN), 2.46 (m, 2H, H-3e-eq and H-3h-eq), 3.79, 3.81 (2s, 6H, 2MeO), 5.62 (m, 2H, H-8e and H-8h), 6.44 (d, 1H, $J_{1,2} = 3.8$ Hz, H-1a), 7.04-8.19 (m, 30H, 6Ph), and 8.61 (s, 1H, C=NH).

Anal. Calcd for $C_{142}H_{162}N_5O_{69}Cl_3$ (3149.2): C, 54.16; H, 5.19, N, 2.22. Found: C, 54.13; H, 4.89; N, 2.14.

 $O - (Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glyc-ero-\alpha-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O - (2,4,6-tri-O-benzoyl \beta-D-galactopyranosyl)-(1 \rightarrow 4)-O - (2-acetamido-3,6-di-O-acetyl-2-deoxy-\beta-D-glucopyranosyl)-(1 \rightarrow 3)-O - [(methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-\alpha-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O - [(methyl 5-Acetamido-4,7,8,9-tetra-0-acetyl-3,5-dideoxy-D-glycero-\alpha-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O - [(methyl 5-Acetamido-4,7,8,9-tetra-0-acetyl-3,5-dideoxy-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O - [(methyl 5-Acetamido-4,7,8,9-tetra-0-acetyl-3,5-dideoxy-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O - [(methyl 5-Acetamido-4,7,8,9-tetra-0-acetyl-3,5-dideoxy-D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O - [(methyl 5-Acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetra-0-acetamido-4,7,8,9-tetr$ $(2,4,6-\text{tri}-O-\text{benzoyl}-\beta-D-\text{galactopyranosyl})-(1\rightarrow 4)-O-(2-\text{acetamido}-3,6$ di-O-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)]-O-(2,4-di-O-acetyl- β -Dgalactopyranosyl)- $(1 \rightarrow 4)$ -O-(2, 3, 6-tri-O-acetyl- β -D-glucopyranosyl)- $(1 \rightarrow 1)$ -(2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (25). To a solution of 24 (24 mg, 0.008 mmol) and (2S,3R,4E)-2-azido-3-O-benzovl-4octadecene-1,3-diol¹³ (18; 20 mg, 0.046 mmol) in dichloromethane (0.5 mL) were added MS-4Å (AW-300, 0.7 g) and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. Boron trifluoride etherate (5 µL) was added, and the mixture was stirred for 8 h at 0 °C and then filtered. The insoluble material was washed with dichloromethane, and the combined filtrate and washings were washed with M sodium hydrogen carbonate and water, dried (Na₂SO₄) and concentrated. Column chromatography (40:1 dichloromethane-methanol) of the residue on silica gel (10 g) gave **25** (10 mg, 46%) as an amorphous mass: $[\alpha]_D$ -12.0° (*c* 0.3, CHCl₃); ¹H NMR (CDCl₃) δ 0.87 (t, 3H, J_{Me,CH2} = 6.6 Hz, MeCH₂), 1.26 (s, 22H, 11CH₂), 1.54, 1.67, 1.78-2.18 (12s, 63H, 17AcO and 4AcN). 2.48 (m, 2H, H-3e-eq and H-3h-eq), 3.79, 3.81 (2s, 6H, 2MeO), 5.60 (m, 2H, H-8e and H-8h), 5.92 (m, 1H, H-5 for sphingosine), and 7.33-8.19 (m, 30H, 6Ph).

Anal. Calcd for C₁₆₅H₁₉₉N₇O₇₁ (3416.4): C, 58.01; H, 5.87, N, 2.87. Found: C, 57.76; H, 5.83; N, 3.16.

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glyc $ero - \alpha - D - galacto - 2 - nonulopyranosylonate) - (2 \rightarrow 3) - O - (2, 4, 6 - tri - O - benzoyl \beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-[(methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$ -O- $(2,4,6-\text{tri}-O-\text{benzoyl}-\beta-D-\text{galactopyranosyl})-(1\rightarrow 4)-O-(2-\text{acetamido}-3,6$ di-O-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)]-O-(2,4-di-O-acetyl- β -Dgalactopyranosyl)- $(1 \rightarrow 4)$ -(2,3,6-tri-O-acetyl- β -D-glucopyranosyl)- $(1 \rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-octadecanamido-4-octadecene-1,3-diol (27). Hydrogen sulfide was bubbled through a stirred solution of 26 (20 mg, 0.006 mmol) in aqueous 83% pyridine (3 mL) for 3 days at 0 °C. The mixture was concentrated, and the residue was stirred with octadecanoic acid (10 mg, 0.035 mmol) and 1-ethyl-3-(3dimethylaminopropyl)carbodiimide hydrochloride(10 mg, 0.053 mmol) in dry dichloromethane (2.0 mL) for 24 h at room temperature. Dichloromethane (50 mL) was added, and the mixture was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (40:1 CH₂Cl₂-MeOH) of the residue on silica gel (10 g) gave 27 (13.2 mg, 62%) as an amorphous mass: $[\alpha]_D$ +9.84° (c 0.26, CHCl₃); ¹H NMR

(CDCl₃) δ 0.89 (t, 6H, 2*Me*CH₂), 1.26 (s, 50H, 25CH₂), 1.54-2.21 (20s, 63H, 17AcO and 4AcN), 2.49 (m, 2H, H-3e-*eq* and H-3h-*eq*), 3.79, 3.81 (2s, 6H, 2MeO), 5.62 (m, 2H, H-8e and H-8h), and 7.22-8.27 (m, 30H, 6Ph).

Anal. Calcd for C₁₈₃H₂₃₅N₅O₇₂ (3656.9): C, 60.11; H, 6.48; N, 1.92. Found: C, 59.91; H, 6.20; N, 1.94

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)- $(2 \rightarrow 3)$ -O- $(\beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O-[(5-acetamido-3, 5-dideoxy-Dglycero- α -D-galacto-2-nonulopyranosylonic acid)- $(2 \rightarrow 3)$ -O- $(\beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)- $(1\rightarrow 6)$]-O-(β -D-galactopyranosyl)-($1\rightarrow 4$)-(β -D-glucopyranosyl)-($1\rightarrow 1$)-(2S,3R,4E)-2-octadecanamido-4-octadecene-1,3-diol (28). To a solution of 27 (13 mg, 0.035 mmol) in methanol (1 mL) was added sodium methoxide (5 mg), and the mixture was stirred for 24 h at 40 °C, and water (0.1 mL) was added. The solution was stirred for 10 h at room temperature, then treated with Amberlite IR-120 (H⁺) resin, and filtered. The resin was washed with methanol, and the combined filtrate and washings were concentrated. Column chromatography (7:40:50 H₂O-MeOH-CHCl₃) of the residue on Sephadex LH-20 (10 g) gave 27 (7.9 mg, quantitative) as an amorphous mass: [α]_D +3.8° (c 0.16, 7:40:50 H₂O-MeOH-CHCl₃); ¹H NMR [98:2 (CD₃)₂SO- $D_2O[\delta 1.01 (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77, 1.81, 1.86, 1.87 (4s, 1.87) (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77, 1.81, 1.86, 1.87 (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77, 1.81, 1.86, 1.87 (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77, 1.81, 1.86, 1.87 (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77, 1.81, 1.86, 1.87 (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77 (t, 6H, 2MeCH_2), 1.21 (s, 50H, 25CH_2), and 1.77 (t, 6H, 2MeCH_2), 1.81 (t, 6H, 2MeCH_2),$ 12H, 4AcN),

Anal. Calcd for C₉₈H₁₇₄N₅O₄₉ (2206.5): C, 53.35; H, 7.95, N, 3.17. Found: C, 53.44; H, 7.82; N, 3.16.

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REFERENCES

- 1. a) S. Hakomori, *Biochem. Biophys. Acta*, **417**, 55 (1975); b) H. Saito, T. Yamagata, and S. Suzuki, *J. Biol. Chem.*, **243**, 1536 (1968); c) N. Sano, S. Yamashiro, and K. Anno, *Biochem. Biophys. Acta*, **343**, 423 (1974).
- 2. a) S. Hakomori, Ann. Rev. Biochem., 50, 733 (1981); b) Glycolipids; New Comprehensive Biochemistry, Vol. 10; H. Wiegandt Ed.; Elsevier, Amsterdam,

1985; c) Ganglioside and Modulation of Neuronal Functions, NATO ASI Series, Series H; Cell Biology Vol. 7; H. Rahman Ed.; Springer-Verlag, Berlin-Heidelberg, 1987; d) S. Tsuji, T. Yamanaka, M. Tanaka, and Y. Nagai, J. Neurochem., 50, 414 (1988); e) G. Walz, A. Aruffo, W. Kolanus, M.Berilaoqua, and B. Seed, Science, 250, 1132 (1990); f) M. J. Polley, M. L. Phillips, E. Wayner, E. Nudelman, A. K. Singnal, S. Hakomori, and J. C. Paulson, Proc. Natl. Acad. Sci. USA, 88, 6224 (1991).

- a) K. P. R. Kartha, A. Kameyama, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., 8, 145 (1989); b) T. Murase, H. Ishida, M. Kiso and A. Hasegawa, Carbohydr. Res., 188, 71 (1989); c) T. Murase, A. Kameyama, K. P. R. Kartha, H. Ishida, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., 8, 265 (1989); d) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., 8, 799 (1989); e) A. Hasegawa, K. Hotta, A. Kameyama, H. Ishida, and M. Kiso, J. Carbohydr. Chem., 10, 439 (1991); f) H. Prabhanjan, H. Ishida, M. Kiso, and A. Hasegawa, Carbohydr. Res., 211, c1 (1991); g) A. Hasegawa, T. Murase, K. Adachi, M. Morita, H. Ishida, and M. Kiso, J. Carbohydr. Chem., 9, 181 (1990).
- a) Fügedi and P. J. Garegg, Carbohydr. Res., 149, c9 (1986); b) M. Ravenscroft, R. M. G. Roberts, and J. G. Tillett, J. Chem. Soc., Perkin Trans. 2, 1569 (1982); c) T. Murase, H. Ishida, M. Kiso, and A. Hasegawa, Carbohydr. Res., 188, 71 (1989).
- a) P. Konradsson, U. E. Uoodong, and B. Fraser-Reid, *Tetrahedron Lett.*, 31, 4313 (1990); b) G. H. Veeneman, S. H. van Leevwen, and J. H. van Boom, *Tetrahedron Lett.*, 31, 1331 (1990).
- a) A. Hasegawa, H. Ohki, T. Nagahama, H. Ishida, and M. Kiso, *Carbohydr. Res.*, 212, 277 (1991);
 b) A. Hasegawa, T. Nagahama, H. Ohki, K. Hotta, H. Ishida, and M. Kiso, *J. Carbohydr. Chem.*, 10, 493 (1991).
- S. Itai, J. Nishikata, N. Takahashi, O. Tanaka, Y. Matsubara, S. Hasegawa, N. Yanai, K. Takaoka, S. Arii, T. Tobe, and R. Kannagi, *Cancer Res.*, 50, 7603 (1990).
- a) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, Carbohydr. Res., 193, c1 (1989); b) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, Carbohydr. Res., 200, 269 (1990).
- 9. Y. Oikawa, T. Tanaka, K. Horita, T. Yoshida, and O. Yonemitsu, *Tetrahedron Lett.*, 25, 5383 (1984).
- 10. R. U. Lemieux, T. Takeda and B. Chung, ACS Symp. Ser., 39, 90 (1976).
- 11. P. J. Garegg, H. Hultberg, and S. Wallin, Carbohydr. Res., 108, 97 (1982).
- 12. K. Jansson, S. Ahlfors, T. Frejd, J. Kihlberg, G. Magnusson, J. Dahmen, G. Noori, and K. Stehvall, J. Org. Chem., 53, 5629 (1988).
- a) R. R. Schmidt and P. Zimmermann, Angew. Chem. Int. Ed. Engl., 25, 725 (1986);
 b) Y. Ito, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., 8, 285 (1989).
- 14. R. R. Schmidt and G. Grundler, Synthesis, 885 (1981).
- 15. T. Adachi, Y. Yamada, I. Inoue, and M. Saneyoshi, Synthesis, 46 (1977).